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# A Novel *NHERF1* Mutation in Human Breast Cancer Inactivates Inhibition by *NHERF1* Protein in EGFR Signaling

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**Abstract.** Background:  $\text{Na}^+/\text{H}^+$  exchanger regulatory factor 1 (*NHERF1*) has been reported to interact with many cancer-related proteins. We recently identified a novel *NHERF1* mutation (E43G) in breast tumours. Materials and Methods: The candidates of *NHERF1* mutation were identified in breast cancer tissues by polymerase chain reaction and DNA sequencing. Wild-type *NHERF1* and E43G mutation were expressed in *NHERF1*-knockdown cells (MCF7 $\Delta$ *NHERF1*) and low-*NHERF1*-expressing cells (SKMES-1). The effects of mutated *NHERF1* on cell functions were examined using *in vitro* methods. Glutathione S-transferase pull-down assays and western blotting were performed to study the effects of *NHERF1* mutation on its interaction with cancer-related proteins. Results: Compared to wild-type *NHERF1*, expression of the mutated *NHERF1* failed to suppress malignant traits in cancer cells, attenuated interaction of *NHERF1* protein with epidermal growth factor receptor (EGFR), and inactivated its inhibition of EGF-induced Akt and extracellular regulated protein kinases (ERK) activation. Conclusion: The results show the causal role of *NHERF1* in the regulation of the EGFR pathway and the progression of breast cancer.

During the past decade, the systemic therapeutic management of breast cancer has undergone a significant transformation. Without targeted therapies, conventional treatment with cytotoxic agents has maximized its potential in terms of patient survival for most types of cancers. Enhanced understanding of the pathogenesis of tumour cell

growth and metastasis has led to the identification of growth signalling pathways as targets for these directed therapies (1). Epidermal growth factor receptor (EGFR) is over-expressed or activated in breast cancer (2, 3), and a number of drugs specifically targeting this tyrosine kinase have been developed over the years as anticancer agents (4, 5). Evidence in cells has shown that the scaffolding protein  $\text{Na}^+/\text{H}^+$  exchanger regulatory factor 1 (*NHERF1*) can associate with EGFR to regulate its trafficking and expression in breast cancer cells and, modulating its role in progression and metastasis of breast cancer (6).

*NHERF1* is a protein with two structural domains named PDZ1 and PDZ2 followed by an ezrin–radixin–moesin-binding region (7, 8). These domains bind to the carboxy-terminal parts of transporters or receptors (9-11). Hence, *NHERF1* binds to the C-terminal of EGFR and stimulation with EGF induces interaction of *NHERF1* with EGFR to regulate localization, degradation and function of the latter (12). *NHERF1* can suppress EGF-induced proliferation of breast cancer cells by inhibiting EGFR phosphorylation and blocking EGFR downstream signalling in breast cancer cells (13).

Loss of heterozygosity (LOH) at the *NHERF1* gene locus (17q25.1) is found in more than 50% of human breast tumours (14), whereas being less frequent in other tumour types, suggesting a pivotal role of *NHERF1* during breast carcinogenesis (14). Previous reports have suggested that *NHERF1* plays a tumour-suppressor role in breast cancer cell lines. *NHERF1* inhibited canonical Wnt signalling and Wnt-dependent cell proliferation in MCF7 and MDA MB-231 cells (15). By interaction with phosphatase and tensin homolog deleted on chromosome 10 (PTEN), *NHERF1* counter-balanced phosphatidylinositol 3-kinase/Akt (PI3K/Akt) oncogenic signaling and enhanced cell responses to platelet-derived growth factor receptor (PDGFR) inhibitor in breast cancer cell lines (16). *NHERF1* knockdown accelerated cell-cycle progression, accompanied by increased expression of cyclin E and elevated retinoblastoma phosphorylation level (14). Furthermore, the mammary glands of *NHERF1*-

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knockout mice exhibit increased mammary duct density accompanied by increased proliferation and  $\beta$ -catenin activity (17). Our previous studies also showed that over-expression of NHERF1 reduced cell proliferation, motility, and invasion of low-NHERF1-expressing SKMES-1 cells, and knockdown of *NHERF1* enhanced the migratory and invasive ability of MCF-7 cells (18). The results of both gain-of-function and loss-of-function studies indicated that NHERF1 acts as a tumour suppressor in tumour cells.

In the present study, a novel *NHERF1* gene mutation (E43G) was identified in a patient with breast cancer. The effects of wild-type (WT) and *NHERF1* mutation E43G on the proliferation, migration, and adhesion of tumour cells were investigated in order to gain insight into NHERF1 function, and to improve our understanding of the role of NHERF1 in tumour development and progression.

## Materials and Methods

**Cell lines and culture.** A human breast cancer cell line, MCF-7 $\Delta$ NHERF1, with low expression of *NHERF1*, was constructed previously (18), by stably expressing a ribozyme targeted to *NHERF1*. Non-small cell lung cancer cell line SKMES-1 was obtained from the European Collection for Animal Cell Culture (Porton Down, Salisbury, UK). *Cos-7* cells were obtained from the American Type Culture Collection (Manassas, VA, USA). All cells were maintained in Dulbecco's modified Eagle's medium (DMEM) supplemented with 10% foetal calf serum, 1% penicillin and streptomycin (Gibco BRC, Paisley, UK) in an incubator at 37°C, with 5% CO<sub>2</sub>, and 95% humidity.

**Human breast specimens.** A total of 20 breast samples were obtained from patients with breast cancer. These tissues were collected immediately after mastectomy, and snap-frozen in liquid nitrogen, with the approvals of the Local Ethical Committee (2013SY15). None of the patients had any underlying medical condition and none were taking any medication.

**Mutational analyses of NHERF.** The genomic DNA from 20 frozen breast cancer tissues was extracted using the DNeasy kit (Qiagen, Hilden, Germany). All six exons of *NHERF1* gene were amplified by PCR using primers corresponding to the neighbouring intronic sequences, and further analyzed by single-strand conformation polymorphism (SSCP) as previously described (19). The sequences of the intronic primers used for the polymerase chain reaction have already been published (20). The candidates of the mutant gene were then identified by DNA sequencing.

**Preparation of expression vectors.** Wild-type glutathione S-transferase (GST)-NHERF1 construct was kindly provided by Dr. Jiale Dai (MD Anderson Cancer Center, Houston, TX, USA). GST-NHERF1 mutation construct was generated from the wild type with the use of Fast MultiSite Mutagenesis System kit (TransGen, Beijing, China). cDNA of *NHERF1*-WT and mutation were cloned into pEASY<sup>TM</sup>-M2 vector using pEASY-Blunt M2 Expression Kit (TransGen) according to the manufacturer's instructions. All constructs were verified by sequencing.

**Cell transfection.** Subconfluent cells were transfected with *NHERF1* expression constructs using lipofectamine reagent according to the manufacturer's instructions (Invitrogen, Waltham, MA, USA). MCF-7 $\Delta$ NHERF1 and SKMES-1 cells were selected in culture medium containing 600  $\mu$ g/ml G418 for 21 days to generate stable *NHERF1*-expressing cell lines. NHERF1 expression was verified by western blotting.

**Wound-healing assay.** The migratory properties of cells were assessed using a wound-healing assay. A variety of cells were seeded at a density of 2 $\times$ 10<sup>5</sup> cells/well into a 24-well plate and allowed to reach confluence. The layer of cells was then scraped with a fine-gauge needle to create a wound of approximately 1500  $\mu$ m. Images of the wound were recorded under a phase-contrast microscope at different time points (0 h, 6 h, 12 h, 24 h). Wound closure/cell migration was evaluated with Image-Pro plus (National Institutes of Health, Maryland, USA).

**GST pull-down assays and western blotting.** GST pull-down assays and western blotting was performed as described previously (15). Anti-NHERF1 was purchased from Becton Dickinson Company (Franklin Lake, New Jersey, USA), anti-extracellular regulated protein kinases (ERK) from Santa Cruz Biotechnology (Santa Cruz, CA, USA), anti-phospho-Akt (Ser473), anti-Akt, anti-EGFR, anti-phospho-EGFR (Tyr1173) were obtained from Cell Signaling Technology (Danvers, MA, USA). Anti-phospho-mitogen-activated protein kinase (MAPK)1/2 (ERK 1/2) was from Millipore (Massachusetts, USA).

**Statistical analysis.** The results were assessed using non-paired (two-sided) Student's *t*-test. A *p*-value of less than 0.05 was considered statistically significant. Each assay was repeated at least three times.

## Results

**Identification of the NHERF1E43G mutation in breast cancer.** The coding region and the intron-exon junctions of the *NHERF1* gene were analyzed by PCR-SSCP (Figure 1A) and confirmed by DNA sequencing (Figure 1B). A previously unreported sequence variant (GAG to GGG), that would result in a switch of codon 43 (Glu-Gly), was identified in the first exon of *NHERF1* in a patient with medullary breast carcinoma. The mutation corresponded to a conserved basic residue in the PDZ1 domain (Figure 1C). The alignment of the NHERF1 protein sequence showed that glutamic acid 43 was highly conserved in the NHERF1 PDZ1 domain throughout evolution and in the PDZ1 domain of the other species in NHERF protein family (Tables I and II), suggesting that it plays an important role in the function of this domain. Furthermore this variant was not found in the 1000 genomes database (<http://browser.1000genomes.org>).

**NHERF1E43G mutation impaired tumour-suppressive effects on malignant cell phenotypes.** Previous studies on the tumour-suppressor functions of NHERF1 were carried out by knockdown or over-expression of *NHERF1* in cancer cell lines (14, 17, 21). Our previous studies also showed that *NHERF1*

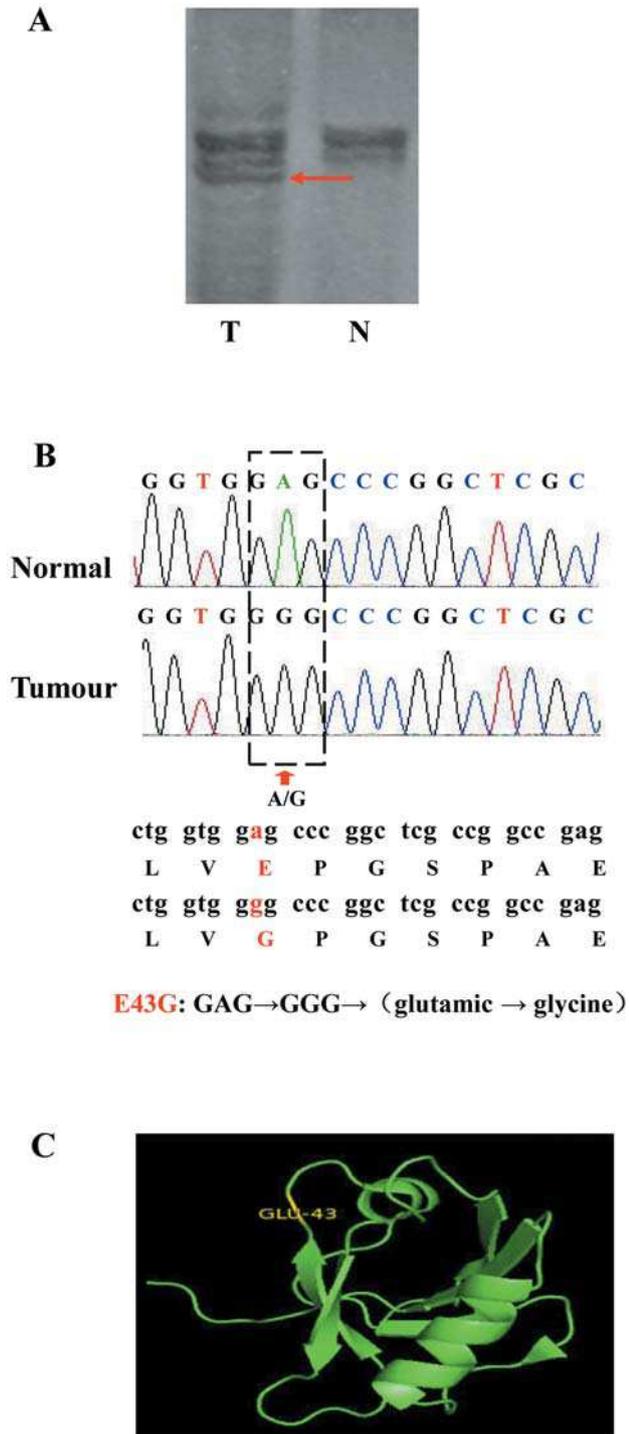


Figure 1. A novel  $\text{Na}^+/\text{H}^+$  exchanger regulatory factor 1 (*NHERF1*) mutation was identified in patients with medullar breast carcinoma. *NHERF1* mutation in exon 1 was analysed by single-strand conformation polymor-phism(SSCP) analysis. A: Abnormal migratory SSCP band patterns for the mutation (T) are demonstrated as compared with normal (N). B: A previously unreported sequence variant (GAG to GGG) was identified by sequencing, which would result in a switch of codon 43 (Glu-Gly). C: The mutation corresponded to a conserved basic residue in the PDZ1 domain.

behaves as a tumour suppressor in breast cancer cells (18). To examine the effects of the E43G mutation on the tumour-suppressive function of *NHERF1*, it was transfected into the low-*NHERF1*-expressing cancer cells SKMES-1 and MCF-7 $\Delta\text{NHERF1}$ . Its expression was assessed by western blotting. The expression level of *NHERF1* was considerably increased in the stably-transfected cells compared to parental and vector control cells, and in cells with E43G mutation was expressed at similar levels to *NHERF1*-WT (Figure 2A and B). Major malignant phenotypes, including cell proliferation and migration, were then evaluated. Compared to vector control cells, cell proliferation was suppressed by up to 22% and 46%, respectively, in SKMES-1 (on day 4) and MCF-7 $\Delta\text{NHERF1}$  (on day 4) cells overexpressing *NHERF1*-WT (Figure 2C and D), cell migration was also reduced by up to 50% and 30% respectively at 24 h (Figure 2E, F). *NHERF1*E43G mutation was able to rescue the cell proliferation and migration that was suppressed by *NHERF1*-WT, since there were no significant differences in proliferation and migration rates between control and the mutant-expressing cells (Figure 2C-F). These results indicate that *NHERF1*-WT acted as a tumour suppressor in SKMES-1 and MCF-7 $\Delta\text{NHERF1}$  cells, and the breast cancer-derived *NHERF1* mutation E43G abolished its suppressor effects on cell proliferation and migration.

*NHERF1*E43G mutation altered interaction of *NHERF1* protein with *EGFR*. It has been shown that *NHERF1* interacts with *EGFR* through its PDZ1 domain and inhibits *EGFR* signalling in breast cancer cells (12, 13, 22). To further investigate if *NHERF1* mutation could affect the interaction between *NHERF1* and *EGFR*, various GST-*NHERF1* fusion protein beads were incubated with protein extracts from COS-7 cells, and *EGFR* was then detected in the pull-down complex. GST-*NHERF1*-WT associated robustly with endogenous *EGFR* protein, whereas GST alone did not. Although *NHERF1*-E43G was able to bind to *EGFR*, its affinity to *EGFR* was reduced by almost 40% compared to that of *NHERF1*-WT (Figure 3). These data indicate that *NHERF1*E43G mutation weakened the interaction of *NHERF1* with *EGFR*.

*NHERF1*E43G mutation inactivated *NHERF1* inhibition of *EGF*-induced Akt and ERK activation. *NHERF1* has been reported to suppress the phosphorylation activation of *EGFR*, blocking *EGFR* downstream signaling in breast cancer cells (13), which in turn controls important biological functions such as cell proliferation and migration (23). Since *NHERF1* mutation affected *NHERF1*-*EGFR* complex formation, it was reasonable to assume that the mutation could also affect *EGFR* signaling. To test this hypothesis, we detected phosphorylation of *EGFR*, Akt and ERK in MCF7 $\Delta\text{NHERF1}$  cells transfected with different *NHERF1* constructs. *EGFR* (Figure 4A), Akt (Figure 4B) and ERK (Figure 4C) were all

Table I. Glutamic acid 43 was highly conserved in the *NHERF1 PDZ1* domain throughout different species.

UniProt Accession Number	Amino acid sequence	Species
O14745	FHLHGEKGGKLGQYIRLV <sup>E</sup> PGSPA <sup>E</sup> KAGLLAGDRLV	HUMAN
P70441	FHLHGEKGGKVGQFIRLV <sup>E</sup> PGSPA <sup>E</sup> KSGLLAGDRLV	MOUSE
Q9JJ19	FHLHGEKGGKVGQFIRLV <sup>E</sup> PGSPA <sup>E</sup> KSGLLAGDRLV	RAT
Q4R6G4	FHLHGEKGGKLGQYIRLV <sup>E</sup> PGSPA <sup>E</sup> KAGLLAGDRLV	MACAQUE
Q3SZK8	FHLHGEKGGKVGQYIRLV <sup>E</sup> PGSPA <sup>E</sup> KSGLLAGDRLV	BOVINE
Q28619	FHLHGEKGGKVGQYIRLV <sup>E</sup> PGSPA <sup>E</sup> KAGLLAGDRLV	RABBIT
Q5ZM14	FHLHGEKGGKPGQYIRLV <sup>E</sup> AGSPA <sup>E</sup> RSGLRAGDRLL	CHICKEN

\*All protein sequences were obtained from UniProt.

Table II. Glutamic acid 43 was highly conserved in the *PDZ1* domain of *NHERF1* and *NHERF2* proteins.

UniProt Accession Number	Amino acid sequence	Protein & Species
O14745	FHLHGEKGGKLGQYIRLV <sup>E</sup> PGSPA <sup>E</sup> KAGLLAGDRLV	NHERF1 HUMAN
Q15599	FHLHGEKGGRRGQFIRRV <sup>E</sup> PGSPA <sup>E</sup> AAAALRAGDRLV	NHERF2 HUMAN
Q9JHL1	FHLHGEKGGRRGQFIRRV <sup>E</sup> PGSPA <sup>E</sup> AAAALRAGDRLV	NHERF2 MOUSE
Q920G2	FHLHGEKGGRRGQFIRRV <sup>E</sup> PGSPA <sup>E</sup> AAAALRAGDRLV	NHERF2 RAT
Q8SQG9	FHLHGEKGGRRGQFIRRV <sup>E</sup> PGSPA <sup>E</sup> AAAALRAGDRLV	NHERF2 RABBIT

\* All protein sequences were obtained from UniProt.

dramatically phosphorylated in both MCF7<sup>ΔNHERF1</sup> parental and vector control cells following 5 min stimulation with 100 ng/ml EGF. NHERF1-WT over-expression significantly inhibited Akt and ERK activation and blocked EGF-induced EGFR phosphorylation. However, the NHERF1 mutation E43G abolished NHERF1-WT inhibition of EGFR, Akt and ERK activation (Figure 4). Meanwhile the base levels of Akt and ERK activation were not detectably different in cells transfected with different *NHERF1* constructs (data not shown). Taken together, these data suggest that cancer-derived *NHERF1* mutation (E43G) abolished the inhibition of the phosphorylational activation of EGFR and its downstream signaling through Akt and ERK signaling pathways induced by EGF.

*NHERF1E43G* mutation resulted in loss of EGF-induced tumour-suppressor effects of *NHERF1*-WT. Since the mutation of *NHERF1* affects EGFR activation in breast cancer cells, *NHERF1* mutation could potentially mediate changes in cell migration of these breast cancer cells. EGF-induced cell migration was investigated in cells stably transfected with *NHERF1*-WT and mutation in order to explore which potential signaling transduction proteins were modulated by *NHERF1*. After pre-culture in serum-free medium for 36 h, cells were treated with 100 ng/ml EGF. NHERF1-WT significantly inhibited the proliferation and migration rate of MCF-7<sup>ΔNHERF1</sup> cells induced by EGF

compared to parental and vector control cells. In contrast, *NHERF1* E43G mutation removed the inhibitory effect of NHERF1 in the EGF-induced malignant cell phenotypes (Figure 5). Taken together, these results suggest that NHERF1 inhibited EGF-induced cell malignant phenotypes in breast cancer cells, but the E43G mutation resulted in complete loss of the tumour-suppressor effects of NHERF1-WT.

## Discussion

Breast cancer is regarded as a heterogeneous group of tumours with different outcomes and responses to treatment. Recent reports suggest a fundamental role for *NHERF1* in human breast cancer. Down-regulation of endogenous NHERF1 in breast cancer cells promoted proliferation, tumorigenic ability, and delayed the turnover of PDGF-induced Akt phosphorylation (24). In our previous study, over-expression of *NHERF1* in a breast cancer cell line suppressed cell proliferation by promoting apoptosis and inhibiting ERK activation (18), suggestive of a tumour-suppressive role for *NHERF1*. However, contrasting results have recently been reported. NHERF1 over-expression in breast tumours is associated with tumour stage, metastatic progression, and poor prognosis (25), implying that NHERF1 is a potential oncoprotein. These contradictory reports suggest a complicated role for NHERF1 in breast cancer.

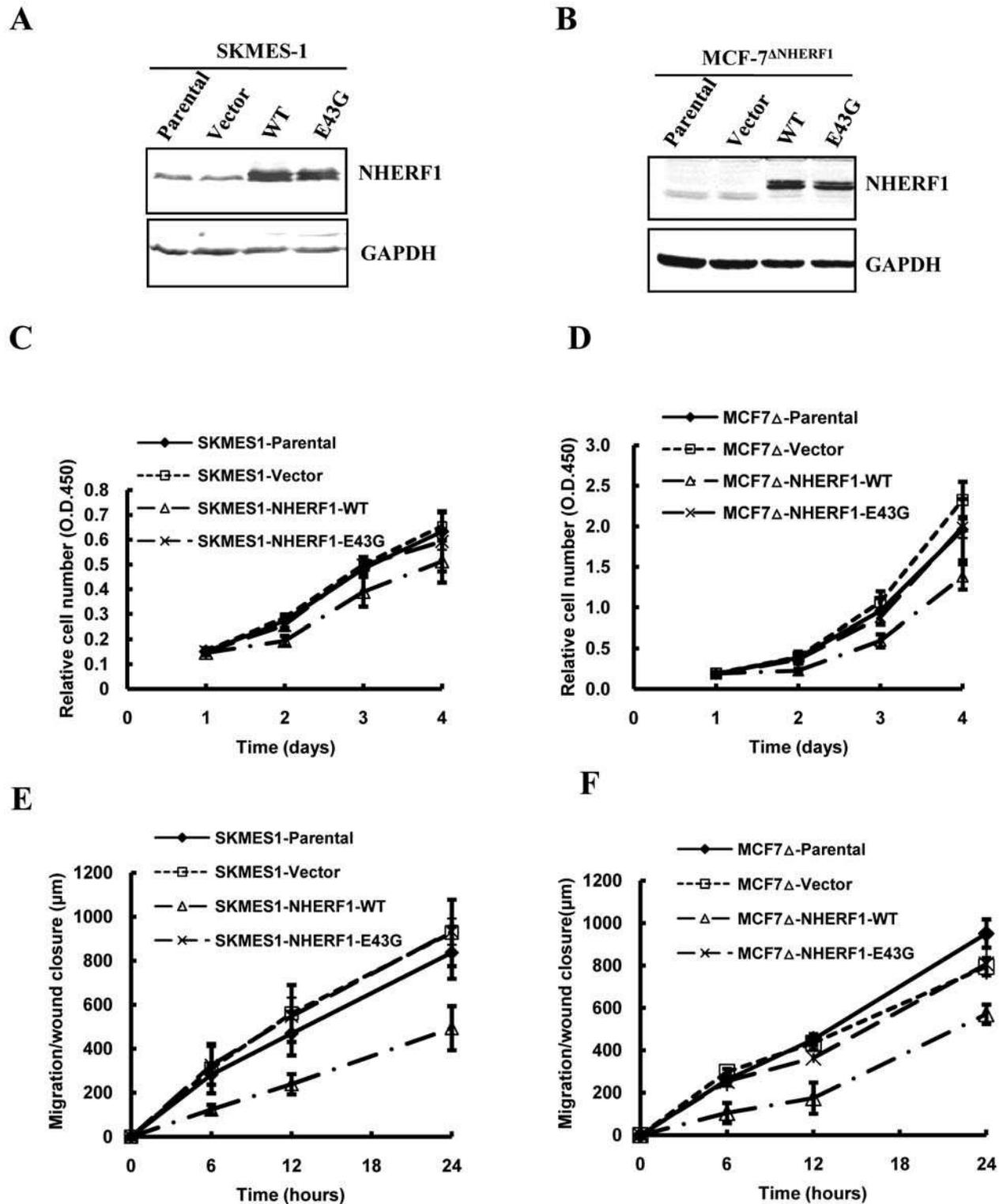


Figure 2. Breast cancer-derived  $\text{Na}^+/\text{H}^+$  exchanger regulatory factor 1 (*NHERF1*) E43G mutation abolished the antitumor effects of *NHERF1* on cell migration in SKMES1 and MCF7 $\Delta$ NHERF1 cells. The wild-type *NHERF1* (WT) and the E43G mutation were stably transfected into SKMES1 and MCF7 $\Delta$ NHERF1 cells and detected with anti-NHERF (A, B). E43G mutation impaired tumor-suppressive function of *NHERF1*, including cell proliferation (C, D) and migration (E, F). The results represent mean values $\pm$ SD of three independent experiments. \* $p < 0.05$ , \*\* $p < 0.01$ . GAPDH: Glyceraldehyde-3-phosphate dehydrogenase.

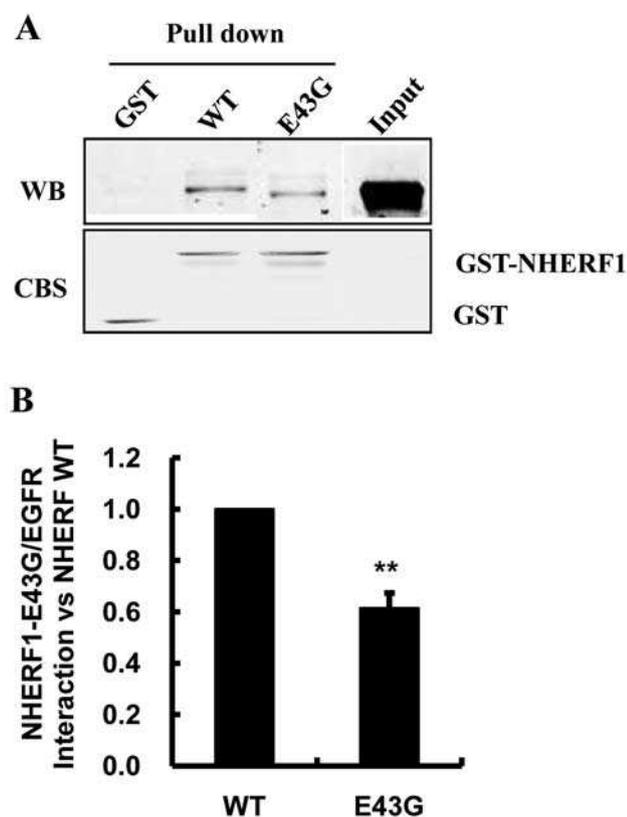


Figure 3. *Na<sup>+</sup>/H<sup>+</sup> exchanger regulatory factor 1 (NHERF1) E43G mutation altered its interaction with epidermal growth factor receptor (EGFR). Glutathione S-transferases (GST)-NHERF1 beads were incubated with COS-7 cell lysates, and then the pull-down complexes were subjected to EGFR immunoblotting. A: NHERF1 mutation (E43G) weakened NHERF1 interaction with EGFR by GST pull-down assay. The Coomassie blue-stained gel was used to visualize the same amount of input of fusion proteins. B: The signals were quantified by densitometric analysis. The results represent mean values $\pm$ SD of three independent experiments. \*\* $p < 0.01$ . WT: Wild-type.*

In this study, the coding region and the intron-exon junctions of the *NHERF1* gene derived from 20 breast cancer tissues were analyzed. A novel *NHERF1* sequence variant (GAG to GGG) resulting in a switch of codon 43 (Glu-Gly) was firstly identified (Figure 1B). The mutation corresponded to a conserved basic residue in the PDZ1 domain (Figure 1C). The sequence is highly conserved according to the alignment of the sequence of PDZ1 of NHERF1 protein in various animals (Tables I and II). These data suggest that a variation in PDZ1 domain could greatly impact the cell phenotype or even cause cell death, making it hard for cells to survive and to identify cells carrying the sequence variation in the PDZ domain. Results of the cell function assays showed that the E43G mutation significantly inactivated the NHERF1 suppression of proliferation and migration of MCF7 $\Delta$ NHERF1

and SKMES-1 cells. These findings further support the tumour-suppressive function of NHERF1 in tumour cells, and suggest that the *NHERF1* mutation may play a causal role in tumour development and progression. Here, we demonstrated that NHERF1 interacted with EGFR (Figure 3) and suppressed EGF-induced proliferation and migration of MCF-7 $\Delta$ NHERF1 cells (Figure 5) by inhibiting EGFR phosphorylation (Figure 4A) and blocking EGFR downstream signaling in breast cancer cells (Figure 4B and C). While *NHERF1* E43G mutation attenuated the interaction of NHERF1 protein with EGFR (Figure 3) and markedly inactivated NHERF1-suppressed activation of Akt and ERK upon EGF stimulation. Hence, loss of suppression of ERK and Akt activities in the cells carrying *NHERF1* mutations might explain their increased malignant cell phenotypes.

The EGFR is one of four members of the HER family of receptor tyrosine kinases. Triggering of EGFR with its potential ligands (EGF, transforming growth factor- $\alpha$  or amphiregulin) lead to homo- and heterodimerization of the ErbB receptors, and subsequent autophosphorylation of the tyrosine domain leads to downstream signalling of Ras/MAPK (26), PI(3)K/Akt (27), phospholipase C- protein kinase C (28), and janus kinase/signal transducers and activators of transcription (29) pathways. This in turn results in cell differentiation and migration. NHERF1, as a signaling adaptor, can associate with EGFR to regulate the latter's functional expression and activation in breast cancer cells and, modulating its role in progression and response to treatment.

The C-terminus of EGFR interacts with NHERF1 through its PDZ1 domain (30), where the E43G mutation of *NHERF1* was identified and located in. It could be imagined that the change in the hydrophobic property of the amino acid residues, [e.g. the acidic amino acid residue (Glu) substituted by the non-polar aliphatic amino acid residue (Gly)], could lead to a change of the spatial structure of the protein, resulting in the different biological activities of NHERF1 in cells. In fact, *NHERF1* E43G mutation altered the binding affinity of NHERF1 to EGFR (Figure 3), and led to the loss of its suppression in the EGFR pathway (Figure 4). However, the mutation could be a useful tool for studying the effects of *NHERF1* on the regulation of the EGFR pathway in cells and the progression and metastasis of breast cancer through the EGFR pathway. Moreover, the one-point mutation in the scaffold protein NHERF1 caused the observed changes in the abilities of proliferation and migration of cells, suggesting that when studying the regulation of the EGFR pathway in breast cancer, some scaffold proteins involved in the pathway should be considered.

In conclusion, a novel *NHERF1* gene mutation (E43G) was isolated and identified in breast tumours. Compared to wild-type NHERF1, expression of the mutated NHERF1 failed to suppress malignant properties of human cancer cells. The E43G mutation attenuated the interaction between

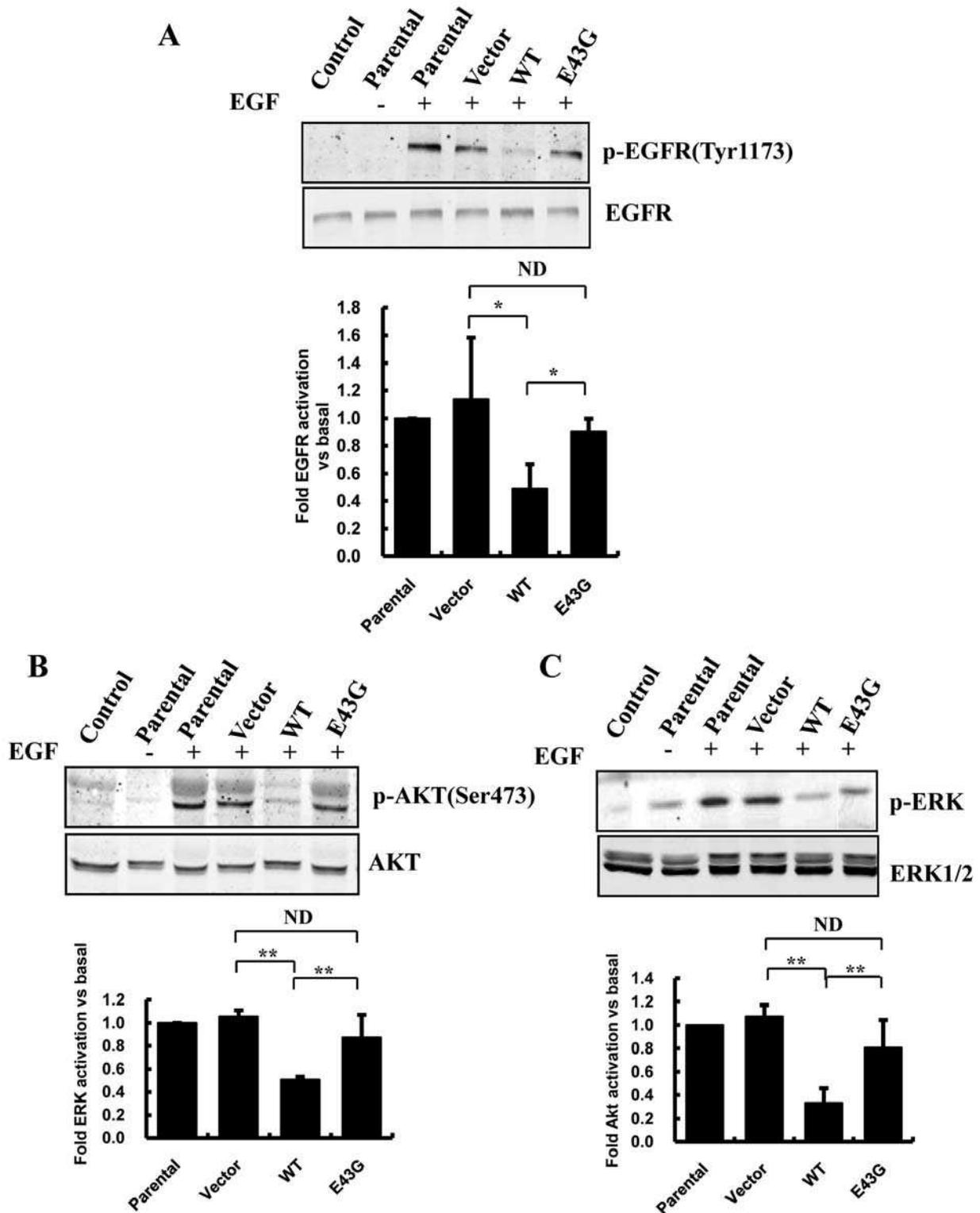


Figure 4.  $Na^+/H^+$  exchanger regulatory factor 1(NHERF1) E43G mutation caused loss-of-function of NHERF1 inhibition of Akt and extracellular regulated protein kinases(ERK) activation induced by epidermal growth factor(EGF). MCF-7 $\Delta$ NHERF1 cells stably transfected with various NHERF1 constructs were serum starved for 36 h and then stimulated with or without EGF for 5 min. Immunoblot was detected with anti-pY1173EGFR (A), anti-EGFR (A), anti-pS473Akt (B), anti-Akt (B), anti-p-ERK (C) and anti-ERK (C). The signals were quantified by densitometric analysis. Data are given as the fold that of stimulated parental cells. The results represent mean values $\pm$ SD of three independent experiments. \* $p < 0.05$ , \*\* $p < 0.01$ . WT: Wild-type.

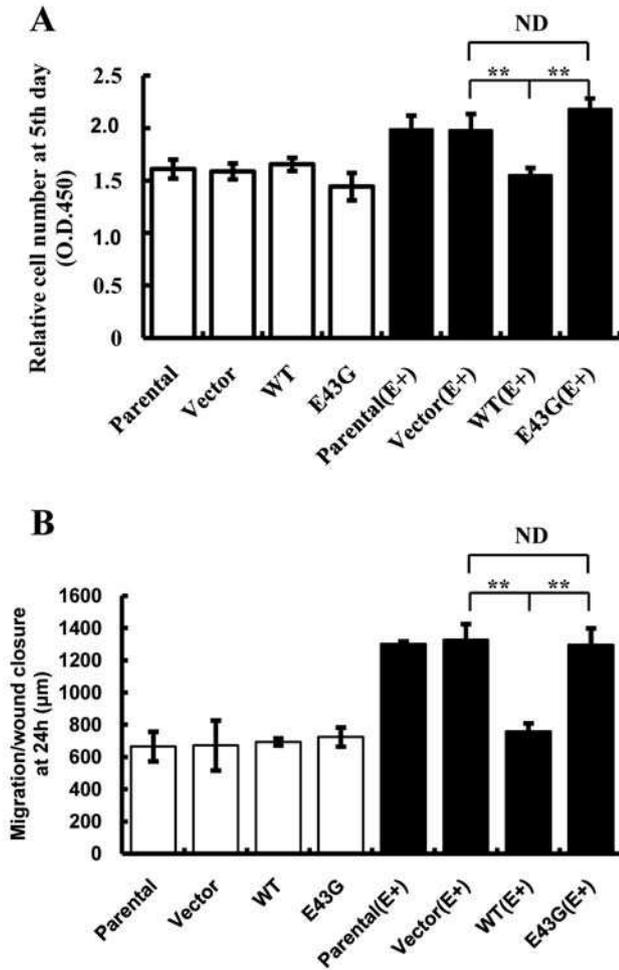


Figure 5.  $Na^+/H^+$  exchanger regulatory factor 1 (*NHERF1*) E43G mutation resulted in loss of the epidermal growth factor (EGF)-induced tumour-suppressive effects of wild-type *NHERF1*-WT in MCF-7 <sup>$\Delta$ NHERF1</sup> cells. The EGF induced cell migration was investigated in cells stably transfected with *NHERF1*-WT and mutation. After pre-culture in serum-free medium for 36 h, cells were treated by EGF (E+). *NHERF1* significantly inhibited proliferation (A) and migration (B) rate of MCF-7 <sup>$\Delta$ NHERF1</sup> cells compared to parental and vector control cells, while *NHERF1* E43G mutation removed the inhibition of *NHERF1* in EGF-induced cell proliferation (A) and migration (B). The signals were quantified by densitometric analysis. The results represent mean values $\pm$ SD of three independent experiments. \* $p < 0.05$ , \*\* $p < 0.01$ . ND: No differences.

*NHERF1* and EGFR proteins, which inactivated *NHERF1* inhibition of EGF-induced Akt and ERK activation, resulting in the total loss of suppression of *NHERF1* in EGF-induced malignant cell phenotypes. These results further demonstrate the functional consequences of breast cancer-derived *NHERF1* mutation (E43G), and suggest the causal role of *NHERF1* in the regulation of the EGFR signaling pathway and the progression of breast tumours.

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