

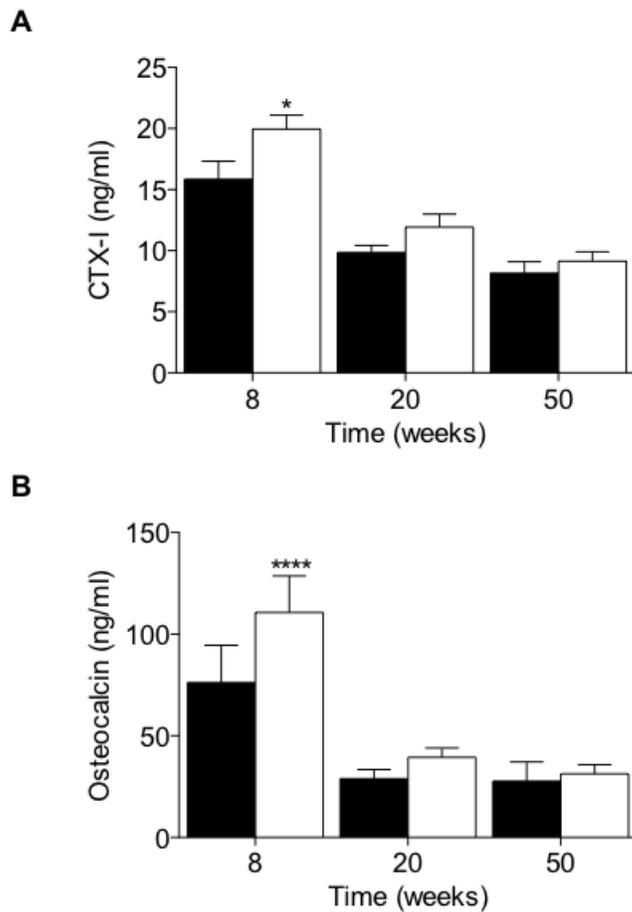
SUPPLEMENTARY MATERIAL

Supplementary Caption:

Supplementary Figure S1: Increased plasma levels of CTX-I (A) and osteocalcin (B) in young male CD59a-deficient mice.

Supplementary Figure S2: CD59a is expressed by murine osteoclasts (OC) and osteoblasts (OB).

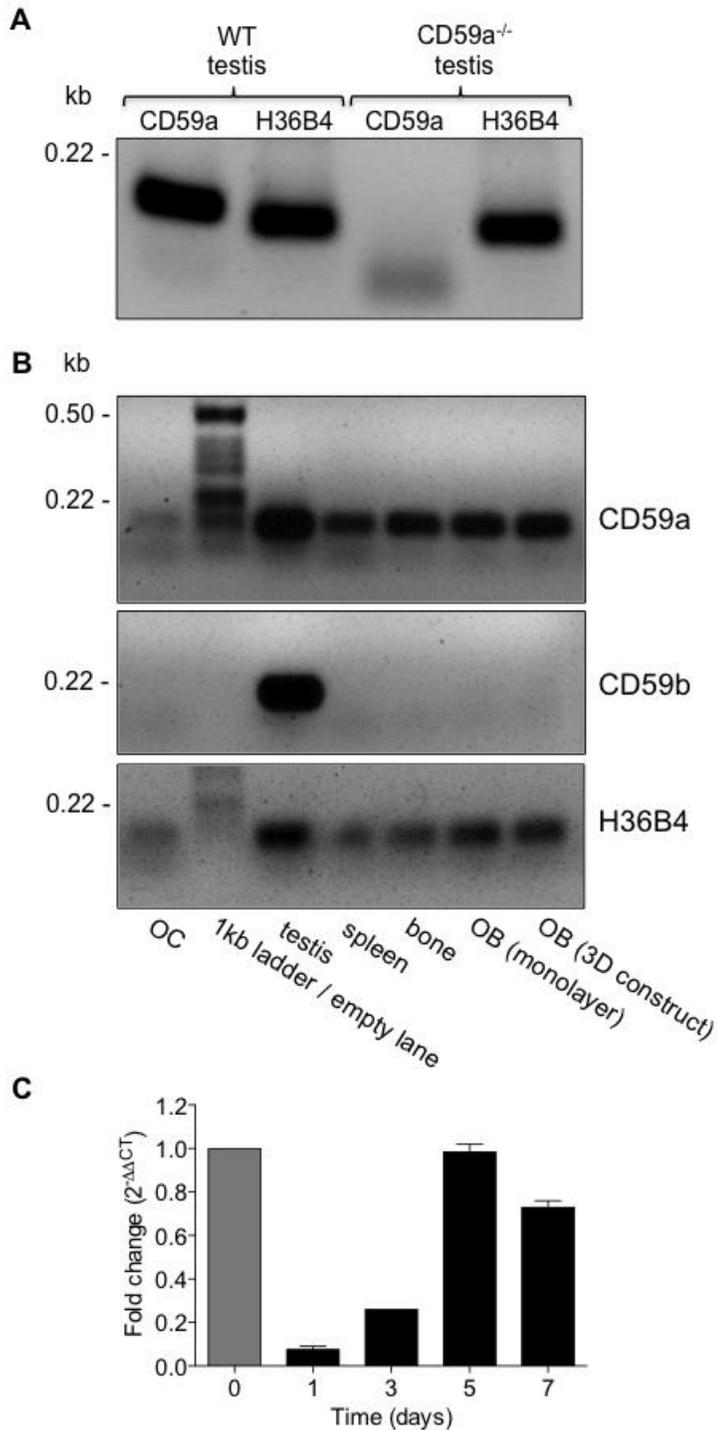
Supplementary Figure S1



Supplementary Figure S1. Increased plasma levels of CTX-I and osteocalcin in young male CD59a-deficient mice. Blood was collected by cardiac puncture in heparin and plasma was obtained after centrifugation. Quantification of murine CTX-I (A) and osteocalcin (B) was performed by ELISA in WT (black bars) and CD59a^{-/-} (white bars)

*samples at 8-10, 20 and 50 weeks of age. ELISAs were performed according to manufacturer's instructions (Immunodiagnostic Systems and Immunotopics Inc. respectively). All values are mean \pm SEM from six separate mice per group. * $P < 0.05$; **** $P < 0.0001$ versus WT levels.*

Supplementary Figure S2



Supplementary Figure S2. *CD59a* is expressed by murine osteoclasts (OC) and osteoblasts (OB). (A) RT-PCR performed with poly(A⁺)-RNA obtained from testis of WT and *CD59a*^{-/-} mice demonstrates specificity of *CD59a* amplification. *CD59a* and *CD59b* are known to be abundantly expressed in germ cells in testis [1]. PCR products were

analysed in 1% agarose gels alongside a housekeeping gene (*H36B4*; [2]) as a positive control. Whereas *H36B4* was amplified in WT and *CD59a*^{-/-} mice, *CD59a* was only detected in WT samples. (B) Expression of *CD59a* in WT OC and OB was confirmed by RT-PCR. For this, poly(A⁺)-RNA was isolated from bone marrow cells differentiated with M-CSF and RANKL for 7 days to differentiate OC, bone tissue (femur), and calvarial OB maintained in monolayer culture or as a 3D construct to obtain an osteocyte phenotype, and analysed as in A. Unlike *CD59a*, *CD59b* expression was reported to be restricted to testicular germ cell elements [1], hence testis and spleen were included in the analysis as a control. *CD59b* was only detected in the testis in contrast to *CD59a* which was expressed in all cells/tissues analysed, including OC and osteoblasts/osteocytes derived through endochondral and intramembraneous bone formation. (C) *CD59a* expression correlates with osteoclast development in differentiation cultures. To further analyse *CD59a* expression during osteoclastogenesis, quantitative PCR was performed for the 7-day time course of differentiation with M-CSF and RANKL. Expression was determined using poly(A⁺)-RNA and a TaqMan assay and is given as change relative to day 0 and normalized for housekeeping gene expression. As expected, levels of *CD59a* expression were high in bone marrow cell isolate (day 0; grey bar). Myeloid cells isolated by differential adhesion had low *CD59a* expression (day 1; black bar) but expression increased ~10-fold upon OC differentiation (day 5-7). Maximal induction between days 3 to 5 is consistent with what is seen for OC markers including cathepsin K [3].

SUPPLEMENTARY REFERENCES

- [1] Harris CL, Hanna SM, Mizuno M, Holt DS, Marchbank KJ, Morgan BP. Characterization of the mouse analogues of CD59 using novel monoclonal antibodies: tissue distribution and functional comparison. *Immunology* 2003;109: 117-26.
- [2] Wagener R, Kobbe B, Aszodi A, Aeschlimann D, Paulsson M. Characterization of the mouse matrilin-4 gene: a 5' antiparallel overlap with the gene encoding the transcription factor RBP-I. *Genomics* 2001;76: 89-98.
- [3] Chen L, Wei XQ, Evans B, Jiang W, Aeschlimann D. IL-23 promotes osteoclast formation by up-regulation of receptor activator of NF-kappaB (RANK) expression in myeloid precursor cells. *Eur J Immunol* 2008;38: 2845-54.