Caffeine protects against experimental acute pancreatitis by inhibition of inositol 1,4,5-trisphosphate receptor-mediated Ca\(^{2+}\) release

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ABSTRACT

Objective Caffeine reduces toxic Ca\(^{2+}\) signals in pancreatic acinar cells via inhibition of inositol 1,4,5-trisphosphate receptor (IP\(_3\)R)-mediated signalling, but effects of other xanthines have not been evaluated, nor effects of xanthines on experimental acute pancreatitis (AP). We have determined effects of caffeine and its xantine metabolites on pancreatic acinar IP\(_3\)R-mediated Ca\(^{2+}\) signalling and experimental AP.

Design Isolated pancreatic acinar cells were exposed to secretagogues, uncaged IP\(_3\) or toxins that induce AP and effects of xanthines, non-xanthine phosphodiesterase (PDE) inhibitors and cyclic adenosine monophosphate (cAMP/cGMP) determined. The intracellular cytosolic calcium concentration ([Ca\(^{2+}\)]\(_{cyt}\)), mitochondrial depolarisation and necrosis were assessed by confocal microscopy. Effects of xanthines were evaluated in caerulein-induced AP (CER-AP), tauroliothocholic acid 3-sulfate-induced AP (TLCS-AP) or palmitoleic acid plus ethanol-induced AP (fatty acid ethyl ester AP (FAEE-AP)). Serum xanthines were measured by liquid chromatography-mass spectrometry.

Results Caffeine, dimethylxanthines and non-xanthine PDE inhibitors blocked IP\(_3\)-mediated Ca\(^{2+}\) oscillations, while monomethylxanthines had little effect. Caffeine and dimethylxanthines inhibited uncaged IP\(_3\)-induced Ca\(^{2+}\) rises, toxin-induced Ca\(^{2+}\) release, mitochondrial depolarisation and necrotic cell death pathway activation; cAMP/cGMP did not inhibit toxin-induced Ca\(^{2+}\) rises. Caffeine significantly ameliorated CER-AP with most effect at 25 mg/kg (seven injections hourly); paraxanthine or theophylline did not. Caffeine at 25 mg/kg significantly ameliorated TLCS-AP and FAEE-AP. Mean total serum levels of dimethylxanthines and trimethylxanthines peaked at >2 mM with 25 mg/kg caffeine but at <100 \(\mu\)M with 25 mg/kg paraxanthine or theophylline.

Conclusions Caffeine and its dimethylxanthine metabolites reduced pathological IP\(_3\)R-mediated pancreatic acinar Ca\(^{2+}\) signals but only caffeine ameliorated experimental AP. Caffeine is a suitable starting point for medicinal chemistry.

INTRODUCTION

Acute pancreatitis (AP) has an incidence of 30 per 100 000 per annum in the UK, commonly caused by gallstones or alcohol excess. Most cases are
mild, whereas a complicated clinical course occurs in one out of every five patients, resulting in significant morbidity, mortality and financial burden. Over the last two decades, our understanding of pathogenesis has advanced, but there is still no specific therapy despite many randomised trials. The development of treatments for AP is, therefore, a priority, one strategy for which is to follow leads from complementary laboratory and clinical studies, as here.

Intracellular Ca\(^{2+}\) signals control normal secretion from pancreatic acinar cells but can become a critical trigger in pathogenesis. Physiological concentrations of acetylcholine (ACh) and cholecystokinin (CCK) generate repetitive elevations in the cytosolic Ca\(^{2+}\) concentration ([Ca\(^{2+}\)]\(_c\)) within the cellular apical pole that elicit stimulus metabolism coupling to generate ATP from mitochondria and stimulus-secretion coupling to initiate exocytosis. Intermittently, global extension of short-lived signals throughout the cell is necessary for normal signalling contributing to transcription and translation. In contrast, toxins such as bile acids, oxidative and non-oxidative metabolites of ethanol and CCK hyperstimulation each elicit abnormal elevations of [Ca\(^{2+}\)]\(_c\) that are global and sustained. These abnormal elevations induce premature activation of intracellular enzymes, mitochondrial dysfunction, impaired autophagy, vacuolisation and necrosis, all of which contribute to the pathogenesis of AP. Ca\(^{2+}\) chelation prevents zymogen activation and vacuolisation through attenuation of Ca\(^{2+}\) overload in acinar cells in vitro and ameliorates the severity of AP in vivo. Blockage of the Ca\(^{2+}\) release-activated Ca\(^{2+}\) channel, also known as the store-operated Ca\(^{2+}\) entry (SOCE) channel, by Orai1 inhibitor GSK-7975A, reduces Ca\(^{2+}\) overload and necrosis in both mouse and human pancreatic acinar cells and prevents AP in three different mouse models. Genetic deletion or pharmacological inhibition of another SOCE channel, transient receptor potential cation channel 3 (TRPC3), also reduces caerulein-induced SOCE and AP.

Excessive Ca\(^{2+}\) release from intracellular stores occurs predominantly via inositol 1,4,5-trisphosphate receptor (IP₃R) Ca\(^{2+}\) channels. The pancreatic acinar cell expresses all three subtypes of the IP₃R in the apical region, close to the luminal membrane, but IP₃R types 2 and 3 are predominantly responsible for physiological Ca\(^{2+}\) signalling and enzyme secretion.

Caffeine (1,3,7-trimethyxanthine) belongs to the methylxanthine class of small, purine-based planar molecules and has several pharmacological actions, including pronounced actions of the IP₃R in the apical region, close to the luminal membrane, but IP₃R types 2 and 3 are predominantly responsible for physiological Ca\(^{2+}\) signalling and enzyme secretion. Caffeine inhibits Ca\(^{2+}\) release from IP₃R by inhibition of phospholipase C-mediated production of IP₃ or by antagonising IP₃Rs through direct binding and reduction of the open-state probability of IP₃Rs. Contrarily, caffeine activates Ca\(^{2+}\) release from ryanodine receptors (RyRs) by increasing the sensitivity of RyRs to Ca\(^{2+}\) itself as observed in multiple cells, although in pancreatic acinar cells effects on IP₃Rs predominate. The effects of caffeine on IP₃-mediated Ca\(^{2+}\) signalling may be protective in AP since the incidence of AP is inversely proportional to the amount of coffee consumed. Caffeine also inhibits cyclic adenosine monophosphate (cAMP) and cyclic guanosine monophosphate (cGMP) phosphodiesterase (PDE), which degrades cAMP and cGMP to non-cyclic forms; inhibition of PDE reduces tumour necrosis factor and leukotriene synthesis, inhibiting innate immunity. Caffeine is a non-selective inhibitor of adenosine receptors, removing an endogenous brake on neural activity. This stimulant effect of caffeine is the most familiar, but taken to excess may result in caffeine intoxication with major central nervous system hyperstimulation. Degradation of caffeine in the liver forms the dimethylxanthines theophylline (1,3-dimethylxanthine), paraxanthine (1,7-dimethylxanthine) and theobromine (3,7-dimethylxanthine), used variously as drugs with similar actions to those of caffeine, although their actions on IP₃Rs or on other SOCE channels, translational to AP, we sought to determine their actions on toxin-induced, IP₃R-mediated [Ca\(^{2+}\)]\(_c\) changes and cell death in vitro, and in three models of AP in vivo.

**MATERIALS AND METHODS**

**Animals**

Adult male CD1 mice (8–12 weeks old) were housed at 23±2°C under a 12 h light/dark cycle with ad libitum access to standard laboratory chow and water. For in vivo experiments, animals were deprived of food but were allowed access to water from 12 h before the start of the experiments.

**Measurements of Ca\(^{2+}\) responses, mitochondrial membrane potential (ΔΨ\(_m\)) and IP\(_3\) uncaging**

Fresh pancreatic acinar cells were isolated as described. Fluoro-4-AM (3 μM), ci-IP\(_3\)/PM (2 μM) and/or tetramethyl rhodamine methyl ester (TMRM, 37.5 nM) were loaded for 30 min at room temperature. Confocal images were acquired on a Zeiss LSM510 system (Carl Zeiss Jena GmbH, Germany) with a 63× C-Apochromat water immersion objective (NA 1.2). ΔΨ\(_m\) was recorded in the perigranular mitochondrial cell region. IP\(_3\) was uncaged by UV excitation of whole cells (364 nm, 1% power) every three seconds where indicated. All fluorescence measurements were expressed as changes from basal fluorescence (F/F₀ ratio), where F₀ represents initial fluorescence at the start of each experiment.

**In vitro necrosis assays**

For CCK-induced cell death, a time-course propidium iodide (50 μM) necrosis assay was run at 37°C using a POLARStar Omega Plate Reader (BMG Labtech, Germany). Isolated murine pancreatic acinar cells (75 μL) were added to a caffeine solution (75 μL) at selected concentrations or the same volume of physiological saline (for controls) prior to CCK (50 nM) addition.

In TLCS-induced cell injury, an end-point propidium iodide (100 μg/mL) necrosis assay was employed. Cells were incubated with respective test solutions and agitated by rotary inversion for 30 min at 37°C, centrifuged (at 260 g for 2 min), resuspended and transferred to a microplate. Data were calculated as background-subtracted (cell-free blanks) percentage of total death (in 0.02% TritonX). Data were normalised to minimum and maximum fluorescence using the formula (Fₐₘₐₓ/Fₐₘₐₓ − Fₐₘᵢₙ) + 1. All experiments were in triplicate.

**Determination of serum dimethylxanthine and trimethylxanthine levels by liquid chromatography-mass spectrometry**

Serum was analysed on a QTRAP5500 hybrid triple-quadrupole/linear ion trap instrument with TurboIon V Ion source (Applied Biosystems, UK), with inline LC (Ultimate 3000 (ThermoScientific/Dionex, UK)) and Gemini C18, 3 μm, 2.1×100 mm column (Phenomenex, UK). Eluent A comprised
RESULTS

Inhibition of ACh-induced \([Ca^{2+}]_c\) oscillations by caffeine and its dimethylxanthine metabolites

ACh (50 nM) caused \([Ca^{2+}]_c\) oscillations in pancreatic acinar cells that were concentration-dependently inhibited by caffeine at 500 \(\mu M\) to 2 \(nM\) (figure 1Ai, ii); 200 \(\mu M\) caffeine resulted in no significant reduction (data not shown). ACh-induced \([Ca^{2+}]_c\) oscillations were also inhibited by 500 \(\mu M\) theophylline (figure 1Aiii) and 500 \(\mu M\) paraxanthine (figure 1Aiv); all dimethylxanthines inhibited ACh-induced \([Ca^{2+}]_c\) signals in a concentration-dependent manner (figure 1Ae). Theophylline, paraxanthine and theobromine induced significantly more inhibition than caffeine at 500 \(\mu M\), with paraxanthine showing the highest potency. In contrast, 1-methylxanthine and xanthine showed minimal inhibition (see online supplementary figure S1A, B).

Inhibition of IP3-mediated \([Ca^{2+}]_c\) signals by caffeine and its dimethylxanthine metabolites

To investigate whether methylxanthines might directly inhibit IP3-R-mediated \(Ca^{2+}\) elevations, a membrane-permeable caged IP3 analogue, c-i-IP3/PM, was loaded into pancreatic acinar cells. Repetitive uncaging of c-i-IP3/PM caused sustained increases of \([Ca^{2+}]_c\) that were inhibited in a concentration-dependent manner by caffeine (3 and 5 mM) (figure 1Bii, ii). Theophylline and paraxanthine showed similar effects (figure 1Biii). These results suggest that methylxanthines inhibit IP3-R-mediated \([Ca^{2+}]_c\) signals by an action on the IP3R.

Caffeine-induced inhibition of CCK-induced \([Ca^{2+}]_c\) signals, \(\Delta\Psi_m\) loss and cell death

The effects of caffeine on CCK-induced toxic, sustained \([Ca^{2+}]_c\) signals were investigated. An elevated \(Ca^{2+}\) plateau followed hyperstimulation with 10 nM CCK (figure 2A), which was reduced by 27% by 1 mM caffeine (figure 2Ai, ii), and blocked by 10 mM (figure 2Aii), effects reversible upon washout. Similar effects were observed when 10 mM caffeine was applied prior to 10 nM CCK stimulation (see online supplementary figure S2A).

Methylxanthines are PDE inhibitors and simultaneous increases in cAMP and cGMP may synergistically inhibit \([Ca^{2+}]_c\) oscillations induced by ACh. The potential contribution of PDE inhibition to the effects of caffeine on CCK-induced sustained \(Ca^{2+}\) signals was investigated using non-hydrolysable analogues of cAMP and cGMP. Addition of 8-bromo-cAMP/GMP (1 mM) did not affect the CCK-induced \([Ca^{2+}]_c\) plateau, whereas 10 mM caffeine caused complete inhibition (figure 2B), suggesting a mechanism independent of intracellular cyclic nucleotide levels, although both xanthine and non-xanthine PDE inhibitors were found to inhibit ACh-induced \([Ca^{2+}]_c\) oscillations (see online supplementary figure S3A–D).

To test potential effects of caffeine on SOCE, internal \(Ca^{2+}\) stores were depleted under \(Ca^{2+}\)-free conditions using either 10 nM CCK or 2 \(\mu M\) thapsigargin, an inhibitor of the sarco-endoplasmic reticulum calcium transport ATPase (SERCA) and SOCE triggered by reapplication of extracellular \(Ca^{2+}\) (5 nM). Following depletion of internal stores with thapsigargin, caffeine was unable to revert the SOCE-induced \(Ca^{2+}\) plateau (figure 2Ci). When 10 nM CCK was used to deplete internal stores, the sustained SOCE plateau was significantly inhibited by 10 mM caffeine in a reversible manner (figure 2Cii). Following application of both CCK and thapsigargin, caffeine did not reduce the associated SOCE (figure 2Ciii). These data, summarised in figure 2Civ, are consistent with an...
Figure 1. Dimethylxanthine and trimethylxanthines inhibit acetylcholine (ACh)-induced and inositol 1,4,5-trisphosphate receptor (IP₃)-induced Ca²⁺ signals in isolated pancreatic acinar cells. (A) Representative traces of ACh (50 nM) induced Ca²⁺ oscillations that were significantly inhibited by caffeine (CAF), theophylline (TP) and paraxanthine (PX): (i) partial inhibition by CAF at 500 μM, (ii) complete inhibition by CAF at 2 mM, or (iii) TP at 500 μM or (iv) PX at 500 μM. (v) Summary histograms of the inhibitory effects of CAF, TP, PX and theobromine (TB) on ACh-induced Ca²⁺ oscillations at both 500 μM and 2 mM. (B) Representative traces of Ca²⁺ elevations (grey) generated by uncaging of the membrane permeable IP₃ analogue, ci-IP₃/PM (2 μM) that were significantly inhibited by CAF (black): (i) partial inhibition at 3 mM and (ii) complete inhibition at 5 mM. (iii) Summary histograms of inhibitory effects of CAF, TP and PX on IP₃-induced Ca²⁺ elevations at 3 and 5 mM. *p<0.05 vs control group; †p<0.05 vs lower concentration. Traces are averages of >20 cells from at least three repeat experiments. Data normalised from basal fluorescence levels (F/F₀) and are expressed as means±SE in histograms.
CER-AP. The serum levels of caffeine were up to 700 μM at 10 min after four caffeine injections (figure 4B). It peaked at 10 min after seven injections of caffeine at >1 mM and gradually reduced to >600 and >400 μM at 2 and 6 h after last caffeine injection, respectively (figure 4B). Caffeine was the most abundant xanthine detected (≈1200 μM 10 min after seven injections), followed by theobromine (≈400 μM), theophylline (≈300 μM) and paraxanthine (≈150 μM) (figure 4C). The total level of dimethylxanthine and trimethylxanthine rose to >2 mM, a concentration capable of exerting marked inhibition of CCK-induced Ca²⁺ signals and cell death.

Effects of dimethylxanthine and trimethylxanthine on the severity of CER-AP
Since caffeine and its dimethylxanthine metabolites were able to protect against Ca²⁺-induced toxicity in vitro, an evaluation of caffeine was carried out in vivo on CER-AP. In the CER-AP with seven caerulein injections, at 12 h after the first caerulein injection, caffeine significantly inhibited necrotic cell death pathway activation (PI uptake) induced by CCK (50 nM) in a dose-dependent manner at 2 and 5 mM (*p<0.05 vs control group; †p<0.05 vs CCK only). Traces are averages of >19 cells from at least three repeat experiments. Data normalised from basal fluorescence levels (F/F₀) and are expressed as means±SE in histograms.

Injection there were significant elevations of serum amylase, pancreatic oedema (pancreatic wet to dry ratio), trypsin and myeloperoxidase (MPO) activity (a marker of neutrophil infiltration), with increases of lung MPO activity, alveolar membrane thickening and serum interleukin (IL)-6 levels (Figure S5, Supplementary Materials and Methods). In CER-AP, but not in CAEE-AP, we assessed renal pathology in CER-AP and in CER-AP following caerulein-induced pancreatic injury, theophylline and paraxanthine did not exert significant protective effects in CER-AP with both 10 and 25 mg/kg regimens (see online supplementary figure S5A–E). To further explore these unexpected findings, the serum levels of theophylline and paraxanthine were measured from both dose regimens. Serum levels of theophylline and paraxanthine 10 min after the last xanthine injection were each <100 μM in the 25 mg/kg regimen and <50 μM in the 10 mg/kg regimen (see online supplementary figure S6). These dimethylxanthine concentrations were previously shown not to alter IP₃R-mediated [Ca²⁺]ᵢ signals in vitro, consistent with an effect of caffeine on this signalling pathway. Since caffeine treatment was markedly protective in CER-AP at 12 h after induction by seven caerulein injections, its effects on more severe disease at a later time point were compared (Figure 6). CER-AP induced by 12 hourly caerulein injections converted mild necrotising AP into a severe necrotising form characterised by extensive pancreatic oedema, neutrophil infiltration and necrosis at 24 h after induction (Figure 6Ei–iv). Caffeine (25 mg/kg regimen) markedly reduced all parameters of pancreatic injury in both models.

**Protective effects of caffeine on TLCS-AP and FAEE-AP**

TLCS-AP caused dramatic increases of pancreatic and systemic injury markers compared with the sham group at 24 h (Figure 7A–E), with marked histopathological changes (Figure 7F). Since pancreatic trypsin activity peaks very early after induction of AP in the bile acid-induced model, this parameter was not included for severity assessment. Caffeine significantly reduced serum amylase (Figure 7A), pancreatic oedema (Figure 7B), and numerous other injury parameters. When compared with the equivalent dose regimens of theophylline (TP) and paraxanthine (PX), caffeine exerted significant protective effects at both 10 and 25 mg/kg (Figure 7G–H).

**Figure 3** Effects of methylxanthines on taurocholic acid 3-sulfate (TLCS)-induced Ca²⁺ signals and cell death. (A) Representative traces showing that the TLCS-induced (500 μM) Ca²⁺ plateau was significantly inhibited by caffeine (CAF): (i) partial inhibition at 3 mM, (ii) the sustained Ca²⁺ plateau was converted to oscillations at 5 mM and (iii) complete inhibition at 10 mM. (B) CAF significantly inhibited necrotic cell death pathway activation (PI uptake) induced by TLCS (500 μM) in a dose-dependent manner at 5 and 10 mM. Similar effects were also seen for (i) theophylline (TP) and (ii) paraxanthine (PX). CAF, TP and PX did not affect basal PI uptake compared with normal controls (*p<0.05 vs control group; †p<0.05 vs TLCS only). Traces are averages of >20 cells from at least three repeat experiments. Data normalised from basal fluorescence levels (F/F₀) for Ca²⁺ signals and from maximal fluorescence levels (F/Fₘₐₓ) for PI uptake, respectively. Data are expressed as means±SE in histograms.
pancreatic MPO activity (figure 7C) and serum IL-6 (figure 7E), but did not affect lung MPO activity (figure 7D). Caffeine significantly reduced the overall histopathological score (figure 7Gi), as well as the specific oedema (figure 7Gii) and inflammation scores (figure 7Giii), with a trend to curtail the necrosis score (figure 7Giv).

Since caffeine inhibits FAEE-induced Ca\(^{2+}\) signals in vitro,\(^7\) its effects in FAEE-AP were tested. Co-administration of ethanol and POA caused typical AP features compared with ethanol alone (figure 8A–G).\(^7\) Two injections of 25 mg/kg caffeine significantly reduced serum amylase, pancreatic oedema, trypsin and MPO activity, although an increase in lung MPO activity was observed (figure 8A–E). The overall histopathological score (figure 8Gi) was greatly ameliorated, with significantly lowered oedema (figure 8Gii) and inflammation (figure 8Giii) with a trend towards a decrease in necrosis (figure 8Giv).

DISCUSSION

This study defines the inhibitory effects of methylxanthines on IP\(_3\)R-mediated Ca\(^{2+}\) release from the pancreatic acinar endoplasmic reticulum store into the cytosol and their potential application in AP. It has been shown that caffeine inhibits IP\(_3\)R as well as IP\(_3\) production in a concentration-dependent manner.\(^28\) We found that inhibition of IP\(_3\)R-mediated Ca\(^{2+}\) release is attributable at least in part to an action on the IP\(_3\)R, since xanthines inhibited IP\(_3\)R-mediated Ca\(^{2+}\) release elicited by uncaged IP\(_3\). Caffeine, theophylline and paraxanthine prevented physiological Ca\(^{2+}\) signalling and toxic elevations of [Ca\(^{2+}\)]\(_{C}\) induced by agents (CCK and TLCS) that cause AP, since xanthines inhibited IP\(_3\)R-mediated Ca\(^{2+}\) release elicited by uncaged IP\(_3\). Caffeine, theophylline and paraxanthine prevented physiological Ca\(^{2+}\) signalling and toxic elevations of [Ca\(^{2+}\)]\(_{C}\) induced by agents (CCK and TLCS) that cause AP, since xanthines inhibited IP\(_3\)R-mediated Ca\(^{2+}\) release elicited by uncaged IP\(_3\). Caffeine, theophylline and paraxanthine prevented physiological Ca\(^{2+}\) signalling and toxic elevations of [Ca\(^{2+}\)]\(_{C}\) induced by agents (CCK and TLCS) that cause AP, since xanthines inhibited IP\(_3\)R-mediated Ca\(^{2+}\) release elicited by uncaged IP\(_3\).
The combined peak concentrations of dimethylxanthine and trimethylxanthine with the 25 mg/kg caffeine regimen were >2 mM, and serum caffeine was >400 μM 6 h after last caffeine injection. Following similar protocols of 25 mg/kg theophylline or paraxanthine, concentrations were far below the effective range on IP3Rs but within the effective range on PDE (approaching 100 μM 10 min after the last dimethylxanthine injection), and no protective effects on in vivo AP were seen. Nor were significant protective effects seen on pancreatic blood flow with the 25 mg/kg caffeine regimen, to be expected if mediated via PDE inhibition. Since pancreatic cellular injury initiates and determines severity in AP, the protective effect of caffeine on AP is likely to have been mediated by inhibition of IP3R-mediated Ca2+ release.

The concentration range over which caffeine inhibited toxic [Ca2+]c overload induced by CCK hyperstimulation was similar to that seen here with quasi-physiological ACh-elicited Ca2+ oscillations, as previously in pancreatic acinar cells and permeabilised vascular smooth muscle cells. There could have been a cAMP/cGMP-dependent component to inhibition of the ACh-elicited Ca2+ oscillations since both xanthine-based and non-xanthine-based PDE inhibitors reduced ACh-elicited Ca2+ oscillations. Nevertheless, PDE inhibition is unlikely to have contributed to the reduction of toxic [Ca2+]c overload as this was not affected by application of cell-permeable cAMP/cGMP analogues, but was immediately reversed upon caffeine administration. It is also unlikely that any increase in SERCA activity occurred in response to caffeine and downstream rises in cyclic nucleotide levels since no decrease in [Ca2+]c was induced by

Figure 5  Dose-dependent protective effects of caffeine on the severity of caerulein acute pancreatitis (CER-AP) at 12 h. Mice received either intraperitoneal injections of 50 μg/kg CER (seven injections hourly) or equal amount of saline injections. Caffeine (CAF) at 1, 5, 10 or 25 mg/kg regimen (seven injections hourly) was begun 2 h after the first injection of CER. Mice were sacrificed at 12 h after disease induction and assessed for (A) serum amylase, (B) pancreatic oedema, (C) pancreatic trypsin activity, (D) pancreatic myeloperoxidase (MPO) activity (normalised to CER group), (E) lung MPO activity (normalised to CER group) and (F) serum interleukin (IL-6). (G) Representative images of pancreatic histopathology for all groups (H&E, ×200). (H) (i) Overall histopathological score and components: (ii) oedema, (iii) inflammation and (iv) necrosis. *p<0.05 vs control group; †p<0.05 vs CER group. Values are means±SE of 6–8 animals per group.

analogues of cAMP and cGMP, which have been shown to upregulate SERCA via phospholamban. Therefore, the actions of caffeine on toxic [Ca\textsuperscript{2+}] overload are consistent with a primary effect on IP\textsubscript{3}R-mediated Ca\textsuperscript{2+} release.

SOCE in pancreatic acinar and ductal cells occurs predominantly via Orai channels and is regulated in part by TRP channels. Previously we found inhibition of Orai to markedly reduce CER-AP, TLCS-AP and FAEE-AP. Inhibition of TRPC3 was found to reduce a mild model of CER-AP while the non-selective cation channel TRPV1 has been implicated in neurogenic inflammation contributing to AP. We obtained no data to indicate any direct effect of caffeine on Orai or TRP channels. On the contrary, SOCE is unlikely to have been inhibited directly by caffeine since caffeine had no

Figure 6 Caffeine (CAF) protects against pancreatic injury in two caerulein acute pancreatitis (CER-AP) models at 24 h. Mice received either intraperitoneal injections of 50 µg/kg CER (both 7 and 12 injections hourly) or equal amounts of saline injections. Caffeine (CAF) at the 25 mg/kg regimen (7 injections hourly) was begun 2 h after the first injection of CER. Mice were sacrificed at 24 h after disease induction and were assessed for (A) serum amylase, (B) pancreatic oedema, (C) pancreatic trypsin activity and (D) pancreatic myeloperoxidase (MPO) activity (normalised to CER group). (E) (i) Overall histopathological score and components: (ii) oedema, (iii) inflammation and (iv) necrosis. *Indicates p<0.05. Values are means ±SE of 6–8 animals per group.

Inhibition of second messenger-mediated Ca\textsuperscript{2+} release via RyR ameliorates both caerulein\textsuperscript{45} and bile acid-induced AP.\textsuperscript{46} Since caffeine enhances Ca\textsuperscript{2+} release from RyRs in excitatory cells,\textsuperscript{32} and RyRs are major contributors to Ca\textsuperscript{2+} signalling in pancreatic acinar cells,\textsuperscript{23,47} the effects of caffeine in the reduction of toxic Ca\textsuperscript{2+} overload observed here might appear contradictory. However, in contrast to the situation in muscle cells, caffeine can only release Ca\textsuperscript{2+} in pancreatic acinar cells under quite exceptional circumstances and then only when present at a low concentration (1 mM); indeed, this effect is abolished by stepping up the caffeine concentration.\textsuperscript{29} Furthermore, ACh-elicited Ca\textsuperscript{2+} signalling is blocked by inhibiting IP\textsubscript{3}Rs pharmacologically\textsuperscript{29} and knockout of the principal subtypes (IP\textsubscript{3}R2 and IP\textsubscript{3}R3) results in a failure of Ca\textsuperscript{2+} signal generation and secretion.\textsuperscript{20} Thus, caffeine is used extensively as an inhibitor of Ca\textsuperscript{2+} release in fundamental investigations of pancreatic acinar and other electrically non-excitable cells.\textsuperscript{27}

Little, if any, protective effect of caffeine on experimental AP can be attributed to actions on adenosine receptors, which have both inhibitory (A1, A3) and excitatory (A2A, A2B) actions mediated in part through changes in cAMP.\textsuperscript{48} Caffeine is an antagonist of all adenosine receptors; the potency of caffeine is highest on A2A then A1 receptors at concentrations 10–20 times lower than on PDE.\textsuperscript{26} In the rat pancreas, few acinar cells express adenosine receptors;\textsuperscript{49} differential subtype expression occurs in vascular endothelium, nerve fibres, islet cells and ductal cells, with total expression A2A>A2B>A3>A1.\textsuperscript{48} While antagonism of the least predominant receptor (A1) previously reduced pancreatic oedema but no other parameter of experimental AP,\textsuperscript{49} the majority of data indicate that increasing adenosine receptor activation by reuptake inhibition or administration

**Figure 7** Protective effects of caffeine (CAF) on tauroliothocoholic acid 3-sulfate (TLCS)-acute pancreatitis (AP). Mice received either retrograde infusion of 50 µL of 3 mM TLCS into the pancreatic duct or underwent sham surgery. CAF at 25 mg/kg (seven injections hourly) was begun 1 h after TLCS infusion. Mice were sacrificed at 24 h after disease induction and were assessed for (A) serum amylase level, (B) pancreatic oedema, (C) pancreatic myeloperoxidase (MPO) activity (normalised to sham group), (D) lung MPO activity (normalised to sham group) and (E) serum interleukin (IL-6). (F) Representative pancreatic histopathology for all groups (H&E, ×200). (G) (i) Overall histopathological score and components: (ii) oedema, (iii) inflammation and (iv) necrosis. *p<0.05 vs other two groups. Values are means±SE of 6–11 animals per group.

of A2 or A3 receptor agonists ameliorates experimental AP.\textsuperscript{50} Furthermore, adenosine receptor activation has broad anti-inflammatory effects, including reduction of neutrophil recruitment and effector functions via A2A and A2B;\textsuperscript{51} antagonism of these receptors may account for the lack of effect of caffeine on lung MPO or lung histopathology in experimental AP. Similarly, protective effects via adenosine receptors would be expected at doses of caffeine that had no (1 mg/kg) or minimal (5 mg/kg) effect.\textsuperscript{52}

High doses of caffeine were required to reduce the severity of experimental AP, with the most effective 25 mg/kg regimen extending into toxicity, indicative of a very narrow therapeutic index. At this dose, the number of hourly injections had to be reduced from seven to two in FAEE-AP to avoid mortality; in CER-AP, 50 mg/kg resulted in caffeine intoxication syndrome, although at 25 mg/kg no visible side effects were observed. In humans, even 10 mg/kg caffeine would be likely to induce caffeine intoxication, with florid neuro-excitotoxic and other undesirable side effects.\textsuperscript{26} The principal caffeine metabolites in humans, monkeys, rabbits, rats and mice are similar and do not differ when given by mouth compared with intraperitoneally.\textsuperscript{39} Paraxanthine, however, is the most abundant dimethylxanthine metabolite in humans, while in mice this is theobromine.\textsuperscript{39}

There is marked individual variability in caffeine metabolism

Figure 8  Protective effects of caffeine (CAF) on fatty acid ethyl ester acute pancreatitis. Mice received two intraperitoneal injections of ethanol (EtOH, 1.35 g/kg) in combination with palmitoleic acid (POA, 150 mg/kg) or equal amounts of EtOH injection only, 1 h apart. CAF at 25 mg/kg (seven injections hourly) was given 1 h after the second injection of EtOH/POA. Mice were sacrificed 24 h after disease induction and assessed for (A) serum amylase level, (B) pancreatic oedema, (C) pancreatic trypsin activity, (D) pancreatic myeloperoxidase (MPO) activity (normalised to EtOH group) and (E) lung MPO activity (normalised to EtOH group). (F) Representative pancreatic histopathology for all groups (H&E, ×200). (G) (i) Overall histopathological score and components: (ii) oedema, (iii) inflammation and (iv) necrosis. *p<0.05 vs other two groups. Values are means±SE of 10 animals per group.
and pharmacokinetics, since the half-life in humans typically ranges from 3 to 7 h, repeated high doses or continuous intravenous infusions would be hazardous unless rapid therapeutic monitoring were to be possible.

Our study has demonstrated proof of principle that caffeine causes marked amelioration of experimental AP largely through inhibition of IP3/R-mediated signalling. Medicinal chemistry starting with the template of caffeine and/or other compounds that inhibit IP3/R-mediated signalling could lead to more potent, selective and safer drug candidates for AP.

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Data sharing statement Upon publication raw data from individual experiments will be made available by the corresponding author to interested researchers requesting data for bona fide scientific purposes.

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